

# 3 mm Drop-in (DI) Packing Instructions

# A. Sample Containers

The sample is contained in a cylindrical sample rotor with the ends sealed by end caps. A pair of DI-4 end caps consists of a front turbine cap and a rear tip cap. To accommodate the many types of samples and experimental conditions, a variety of rotors, end caps, and sealing cells are available. The optimum rotor and caps for a particular experiment depend on the following considerations: (1) sample availability; (2) significance of weak background signals from the caps; (3) temperature range of the experiment; (4) physical properties of the sample (e.g., wet, dry, air sensitive, etc.); (5) required spinning speed; and (6) the importance of optimizing microwave efficiency for DNP.

The DI rotors are manufactured with extremely close tolerances and will work with all the different types of DI end cap pairs for that size. End cap pairs are available in a variety of materials.

Refer to the NMR Probe and Rotors Materials Data in your probe manual or on the Doty Scientific web site for more information on the properties of various rotors and turbine caps.

## B. Sample Loading

A well balanced, uniformly packed sample is required to obtain optimum sample spinning and  $B_0$  field homogeneity. In packing the rotor, symmetry is critical <u>around</u> the spinning axis of the sample. Variations in the packing density along the length of the rotor are not critical. If very high  $B_1$  homogeneity is required, the sample needs to be restricted to the central half of the full sample length using special spacers or cells. A rotor with the sample packed preferentially toward one end will still spin well if the packing is radially symmetric. A well balanced sample can be obtained by hand packing if the material is kept uniform while filling the rotor. In many instances, better packing is achieved with the aid of a bench spinner. Examples of both methods are described below.

# 1. Sample Preparation

Most solids and wet samples can be packed uniformly into the rotors for spinning. Plastics can be machined into solid plugs. Irregular crystals or non-uniform powders are readily ground fine using a mortar and pestle, and fibers can be minced, ground, or even wound in such a way as to be radially symmetric. Spinning of granular samples may be



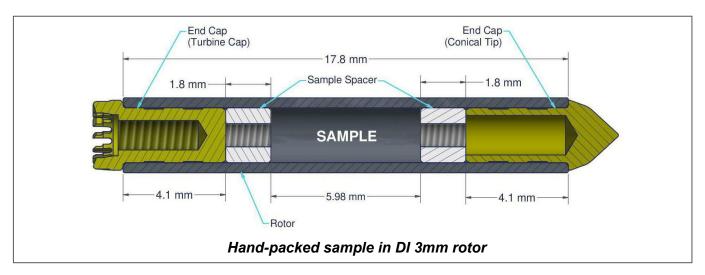
improved by mixing the material with a fine powder lacking interfering NMR signals. Tissues and fibers may spin better when packed wet. Some other materials should be dried and ground fine for best packing. A small funnel cup, a tamp, a rotor scraper, and sample leveling tools are provided to assist in packing rotors.

### 2. Hand Packing

Inspect the rotor for cracks. **Never use cracked rotors**. You should inspect the rotor with a magnifying glass. Sometimes cracks will show up with the following technique. Mark the rotor with a permanent marker. Wipe the color off with a towel wet with alcohol. The marker color may remain in a crack after cleaning. Also check the turbine and tip caps to see that they have not been damaged.

- a. Note: Torlon and GFT caps have some moisture absorption. It may be periodically necessary to bake out the turbine caps or tip caps at 50°C for one hour. This is necessary if the caps become too tight. (The opposite condition is much less likely. However, if one is in a very arid area or operating in a low moisture environment, the caps may have to be soaked in a liquid too make them tighter.) Glass Filled Torlon (GFT)caps are used for extended temperatures.
- **b.** Insertion of the <u>Conical Tip Cap</u>: The proper cap insertion tools should be used to avoid damage to the conical tip cap. Gently insert a conical tip cap partly into one end (the painted end if the rotor is painted) of the rotor. Then place the rotor with the tip cap into the conical tip insertion tool. Place the top insertion tool

- on the other end of the rotor. Press the rotor between the two insertion tools to fully seat the tip cap. The cap must be pressed in completely and squarely. (There should not be a gap between the cap head and the rotor.)
- **c.** *Optional...* Kel-F, PPS and Teflon sample spacers are available to restrict sample. (Teflon spacers are made tighter so that they can be used for liquids or solids.) Insert a sample spacer as shown in the figure below. Ensure that the spacer is placed such that it seats squarely against the inner end of the conical tip cap Check the conical tip again to ensure it is still seated as described above.



- **d.** Using the plastic funnel and spatula, fill the rotor about 30% full with the sample. It is critical that the sample be uniform. Loading of sensitive materials can be performed in a glove box, under special lighting, etc. With the tip cap in the conical tip insertion tool, lightly tap the bottom of the insertion tool squarely against a smooth, horizontal surface to cause the sample to settle.
- e. The sample may now be packed more tightly by placing the tamping tool in the top of the rotor (which is still being held in the bottom insertion tool) and again gently tap.
  Note: Using the tamp increases the concentration of the sample and therefore the S/N ratio, but it will also tend to accentuate imbalance in granular samples, or samples with irregular particle sizes.
- f. Repeat steps 4 through 6 to fill the rotor, and then level the sample with the sample leveler.
- **g.** Optional: Insert the second Sample Spacer as shown above. Take care that the threaded end faces outwards, necessary for ease of removal with the tool provided, and to ensure the sample stays uniformly packed.
- h. Alternative Packing Tips. For best homogeneity, samples of limited quantity may be packed with spacers to confine the sample to the center of the sample coil as described above. Or they may be packed simply by partially filling the rotor and spinning. Centrifugal force will usually pack the sample against the wall in a well-balanced distribution. (Rotor spin packing is also useful with hydroscopic samples. Fill, spin, fill the vortex, spin... and repeat.)
- i. Insertion and removal of DI-3 turbine caps is presented in the table in section D on the next page.

  Note: If you experience difficulty in cap removal, the caps can be more easily removed by cooling below -80°C. This can be done by placing the rotor in dry ice or liquid nitrogen for a few minutes. Remove the rotor from the cold bath, using a cloth or glove to protect the fingers, and immediately remove the turbine cap with the technique in D-2a while the rotor is still cold. Then the sample can be removed from the rotor. After the sample has been removed, the tip cap can then be removed as described in D-2b

#### C. Sample Removal

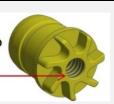
After the experiment, the turbine cap can be removed by the method in **D-2a** on the following page. The sample may be scraped out with the scraper or, in stubborn cases, dissolved. Standard turbine caps are designed to fit very tightly when new. They will loosen slightly with each use, especially if used much above room temperature.

Alumina, Kel-f, and Kalrez may be cleaned with virtually anything except very hot, concentrated, strong bases and Hydro-fluoric acid. Zirconia, macor, silicon nitride, Viton, Vespel, Aurum, and PEEK may be cleaned with most organic solvents, and cold, moderately concentrated (40%) strong acids and bases for several minutes. Hot soapy water often works best on the ceramics. Viton and Kalrez swell in acetone and some other organics. Torlon, GFT, and Vespel swell in water.

# D. Insertion and Removal of DI-3 Turbine Caps

# **Current Design DI-3 Turbine Caps**

Note how the turbine blades are connected to form a ring around the threaded hole.



# Proper Insertion Tools for DI-3 Turbine and Tip Caps



# 1. Current Design Caps – Proper Turbine Insertion:

a. For the DI-3 turbines, the proper cap insertion tools (shown above) should be used to avoid damage to the turbine or the conical tip cap. After filling the rotor (see prior sections), the rotor with the tip cap should be sitting in the conical insertion tool. Gently insert the turbine part way into the rotor. Now place the top insertion tool over the turbine cap and push the cap into the rotor.

Note: Kel-f turbine caps are softer than Torlon and GFT. Kel-F turbines should be inserted (and removed) using the brass removal tool shown below rather than the top insertion tool shown above.

**b.** If the cap does not fully seat (there should not be a gap between the cap head and the rotor), some sample will need to be removed using the sample leveling tool. Remove the cap. Remove some sample with the leveling tool and reinsert the cap using the process described above.





### **Proper Removal Tools for Current Design Caps**

### 1. Proper DI-3 Cap Removal Technique.

**a.** For turbine removal, the rotor should be held in the rotor holder (shown above). The rotor holder is easier to hold than the rotor, and protects the rotor while the cap is pulled out. The rotor is inserted into the rotor holder with the turbine end first. (The rotor can only fit into the holder

at its larger end.) The puller can then be threaded into the turbine. The turbine can be pulled out through the smaller end of the holder. It is important to pull the turbine straight out (not at an angle).

**b.** With the front turbine cap removed <u>and the rotor emptied</u> of all sample, the rear tip cap can be removed using the rotor holder and plunger. The rotor is inserted, (with the remaining rear tip end first) into the rotor holder. The plunger is inserted into the rotor. The tip cap is pressed out – passing through the smaller end of the rotor holder.



When caps are new it may be necessary to cool them in liquid nitrogen to get them out. They may be tight at first. Note section **B-2h** in Hand Packing on the prior page.

There is a simple video on the use of packing tools. Note: In the video, JB mentions packing the sample, then removing the cap to see if there is a significant hole in the center of the rotor. He then he instructs filling the rotor more completely. This should only be needed when maximum sample is required. (Most of the time the sample is fine as first packed.) To access the video, <a href="https://dotynmr.com/packing-video">https://dotynmr.com/packing-video</a>.